



## Effect of sodium para nitrophenolate 0.3% SL on growth and yield of chilli (*Capsicum annuum L.*) in Inceptisols

S. K. DAS, \*S. SAMUI AND P. NANDY

Department of Agronomy, Faculty of Agriculture, Bidhan Chandra Krishi Viswavidyalaya, Mohanpur, Nadia, West Bengal-741252, India.

Received : 04.05.2022 ; Revised : 23.09.2022 ; Accepted : 30.09.2022

DOI : <https://doi.org/10.22271/09746315.2022.v18.i3.1621>

### ABSTRACT

Plant growth regulator (PGR) is a non-nutritive organic substance that can change plant physiological activities at a very low concentration. Field experiments were conducted at Kalyani D Block Farm, Kalyani, Nadia under Bidhan Chandra Krishi Viswavidyalaya, Mohanpur, West Bengal during rabi and kharif seasons of 2017-18 and 2018-19. Six different treatments including sodium para-nitrophenolate (Na-p-NP) 0.3 % SL (at 4, 8 and 16 ml L<sup>-1</sup>), triacontanol (TRIA) 0.1 % EW at 250 ml ha<sup>-1</sup>, alpha naphthyl acetic acid (NAA) 4.5 % SL at 10 ppm and untreated control were evaluated in rabi and kharif chillies (var. Bullet Chilli) in Inceptisol. All the PGRs were applied for three times as foliar application in both the seasons at 25, 45-50 and 65-70 days after transplanting (DAT). The results revealed that the growth attributes (plant height, number of secondary branches and flowers plant<sup>-1</sup>) were significantly improved with the application of Na-p-NP at 16 ml L<sup>-1</sup>. It also improved the yield attributes (number of fruits plant<sup>-1</sup>, fruit weight, length, width) and ultimately the fruit yield during both the seasons. There was no significant effect of PGRs on number of primary branches plant<sup>-1</sup>, days to 50% flowering and maturity of chilli crop.

**Keywords:** Chilli, fruit yield, growth attributes, PGR, yield attributes.

Chilli (*Capsicum annuum L.*) is a very important spice as well as vegetable crop. As chilies are grown in all seasons in many parts of India, the fruits are accessible throughout the year. India is the largest producer, producing 1.75 million tons, and accounting for 43% of world chilli output, followed by China, Ethiopia, Thailand, Pakistan and Bangladesh (Anon., 2021a). India ranks first in terms of export and consumption of chilli. The colour and pungency levels of Indian chilli are regarded to be the world famous commercial features. Chillies from India are mostly supplied to Asian nations such as China, Sri Lanka, Malaysia, Bangladesh, Singapore, Thailand and the United Arab Emirates, among others. Andhra Pradesh comes on top in dry chilli output in the country during 2019-20, with a production record of 0.67 million tons from an area of 0.14 million hectares and productivity of 4.66 t ha<sup>-1</sup>, followed by Telangana, Madhya Pradesh, Karnataka and West Bengal (Anon., 2021b).

Production of chilli is affected by a number of parameters viz. genetics, the environment and cultivation methods. Number of flowers and fruits decreases as a result of a hormonal and physiological abnormality in plants caused by nondurable circumstances, reducing chilli output (Vega-Alfaro *et al.*, 2021). One of the most serious issues with chillies is excessive flower and fruit dropping, which results in a low chilli production. If the drop of flowers is addressed, chilli yield may be boosted. This problem may be overcome through

selecting breeding lines that preserve a high percentage of blooms, or by using physiological treatments such as spraying plant growth regulators (PGRs) to prevent flower loss (TamilSelvi and Vijayaraghavan, 2014).

The PGR is a non-nutritive organic substance that can change plant physiological activities and also becomes effective at a very low concentration (Gianfagna, 1995). They contribute to crop growth, productivity and quality by working within plant cells (Karali and Mohapatra, 2007). They have a complete control over the quantity, direction and type of plant development, resulting in remarkable gains in plant growth and yield (Shah and Ahmad, 2006; Emongor, 2007). Although plants produce their own PGRs, various studies have confirmed that the plants may respond to exogenously applied growth hormones. Externally provided hormones can be stored by plants as reversible conjugates, which can release active hormones as per the needs of the plant throughout the growing season (Tiwari *et al.*, 2011). PGRs can improve the source-sink relationship and increase photo assimilate translocation, resulting in improved fruit set (Anolisa *et al.*, 2020).

Nitrophenolates (NP) are phenolic substances that have a role in the metabolic processes of the plants. For example, they can promote development and growth, boost endogenous auxin levels, increase nutrient absorption by plant roots, and promote antioxidant and photosynthetic activities of plants (Valero *et al.*, 2014). According to the studies on the influence of PGRs on

Email: sumansamui369@gmail.com

How to cite : Das, S.K., Samui, S. and Nandy, P. 2022. Effect of sodium paranitrophenolate 0.3% SL on growth and yield of chilli (*Capsicum annuum L.*) in Inceptisols. *J. Crop and Weed*, 18 (3): 86-91.

vegetable and fruit crops of the Solanaceae family, the use of various PGRs has been found to reduce flower and fruit drop, resulting in increased chilli yield per unit area and per unit time. Sodium para-nitrophenolate 0.3% SL (Na-p-NP 0.3%) is commonly and effectively employed in the production of the world's most significant crops. Its use in the chilli plants improves plant growth and yield (Surendar *et al.*, 2020). Chilli's distinct reactions to various PGRs have also been documented (Joshi *et al.*, 1999; Balraj *et al.*, 2002). Green pepper quality can be maintained throughout harvest and also after harvest by using gibberellic acid (dos Anjos *et al.*, 2022). Foliar application of gibberellic acid (GA), 1-Naphthaleneacetic acid (NAA), and p-Chlorophenoxyacetic acid (CPA) shows the best effect in improving fruit set (Sreenivas *et al.*, 2017; Akhter *et al.*, 2018; Mahindre *et al.*, 2018), fruit yield (Patel *et al.*, 2016; Gare *et al.*, 2017; Tapdiya *et al.*, 2018; Ahmed *et al.*, 2021), and fruit quality in chillies (Deshmukh *et al.*, 2010; Kar *et al.*, 2016; Ahmed *et al.*, 2021). Due to content of amino acids, vitamins and carbohydrates, triacontanol can boost fruit set and production in chillies (Srivastava, 2007).

With these perspectives in view, the current investigation was carried out to assess the bio-efficacy of sodium para-nitrophenolate 0.3% SL (PGR) on chilli in Inceptisols during *rabi* and *kharif* seasons.

## MATERIALS AND METHODS

A field experiment was undertaken under the sub-humid subtropical climatic condition and medium land situation at Kalyani D Block Farm, Bidhan Chandra Krishi Viswavidyalaya, Kalyani, Nadia, West Bengal during *rabi* and *kharif* seasons of 2017-18 and 2018-19. The experimental site was situated in the new alluvial zone (NAZ) of West Bengal, and located at 22°58' N latitude and 88°3'E longitude with an altitude of 9.75 m above mean sea level. The soil was sandy clay loam in texture having a pH of 7.6, and mostly fertile and deep as developed from recent alluvium of the river Ganges. The variety chosen for the study was Bullet Chilli for both the seasons with duration of 144 days during *kharif* and 146 days during *rabi*. Six different treatments viz. sodium para-nitrophenolate (Na-p-NP) 0.3 % SL (4, 8 and 16 ml L<sup>-1</sup>), triacontanol (TRIA) 0.1 % EW (250 ml ha<sup>-1</sup>), alpha naphthyl acetic acid (NAA) 4.5 % SL (10 ppm) and untreated control were assigned in a randomized block design with four replications. Individual plot size was 5 m x 5 m and spacing was 60 cm x 45 cm. All the plots received the recommended doses of fertilizers at 120: 60: 60 kg N: P<sub>2</sub>O<sub>5</sub>: K<sub>2</sub>O ha<sup>-1</sup> in both the seasons. 50% N along with full doses of P<sub>2</sub>O<sub>5</sub> and K<sub>2</sub>O were applied as basal and incorporated into the soil at two days prior to transplanting, and the

rest 50% N was top dressed at 30 days after transplanting (DAT). All the PGRs were sprayed at three times as foliar application in both the seasons, keeping the first, second and third spray at 25, 45-50 and 65-70 DAT, respectively. No pesticides were used in the study. Five plants were randomly selected and tagged from each treatment in different replicates. Observations were recorded from the selected plants only on number of primary and secondary branches plant<sup>-1</sup>, height of the plants (cm), number of flowers plant<sup>-1</sup>, number of fruits plant<sup>-1</sup>, days to 50% flowering and maturity, fruit length (cm), width (cm), weight (g) and fruit yield (t ha<sup>-1</sup>).

## RESULTS AND DISCUSSION

### *Number of branches plant<sup>-1</sup>*

The treatments had a significant influence on the number of secondary branches plant<sup>-1</sup>, whereas it was non-significant on the number of primary branches plant<sup>-1</sup> (Table 1). The highest number of primary and secondary branches plant<sup>-1</sup> was recorded under Na-p-NP at 16 ml L<sup>-1</sup> in both the seasons. Maximum (4.7) and minimum (3.6) number of primary branches plant<sup>-1</sup> were recorded in Na-p-NP at 16 ml L<sup>-1</sup> and TRIA (250 ml ha<sup>-1</sup>), respectively during *rabi* season, whereas it was maximum (8.5) and minimum (5.5) in number for the secondary branches plant<sup>-1</sup> as recorded in Na-p-NP at 16 ml L<sup>-1</sup> and untreated control, respectively. The highest number of primary branches (5.2) and secondary branches (10.5) plant<sup>-1</sup> in *kharif* chilli were observed in Na-p-NP at 16 ml L<sup>-1</sup>, whereas these were the lowest (4.0 and 6.1 for primary and secondary branches plant<sup>-1</sup>, respectively) in untreated control. It might be explained with the rise in the number of primary branches due to auxins activating cell elongation and cell division in the axillary buds. More number of branches was formed as a result of native cytokinin production in root cells and its subsequent transfer to the auxiliary buds. Positive effect of PGRs on branch number plant<sup>-1</sup> was also reported earlier (Chaudhary *et al.*, 2006; Kannan *et al.*, 2009; Kalshyam *et al.*, 2011; Vandana and Verma, 2014).

### *Plant height*

Plant height, an essential morphological factor, was significantly influenced due to different concentrations of Na-p-NP and NAA 10 ppm. Application of Na-p-NP at 16 ml L<sup>-1</sup> recorded the highest plant height (100.6 and 120.1 cm), being about 67.12% and 62.3% more than that of untreated control in *rabi* and *kharif* chilli, respectively. This might be because of the meristematic region's substantial rise in cell elongation and cell division. Na-p-NP was found to have a favourable effect on plant height (Djanaguiraman *et al.*, 2005; Kozak *et al.*, 2008; Surendar *et al.*, 2020) due to the interaction

*Effect of Sodium Para Nitrophenolate 0.3% SL on Growth and Yield of Chilli*

**Table 1: Effect of treatments on growth attributes of rabi and kharif chilli (pooled of two-year data)**

Treatment	Primary branches plant <sup>-1</sup>		Secondary branches plant <sup>-1</sup>		Plant height (cm)		Flowers plant <sup>-1</sup>		Days to 50% flowering		Days to maturity	
	Rabi	Kharif	Rabi	Kharif	Rabi	Kharif	Rabi	Kharif	Rabi	Kharif	Rabi	Kharif
Na-p-NP (4 ml L <sup>-1</sup> )	4.0	4.3	6.2	7.2	81.8	96.5	198.6	210.8	50.8	49.8	145	142.0
Na-p-NP(8 ml L <sup>-1</sup> )	4.3	4.8	7.1	8.1	98.5	115.7	220.8	260.3	53.5	52.6	146	144.0
Na-p-NP(16 ml L <sup>-1</sup> )	4.7	5.2	8.5	10.5	100.6	120.1	238.2	276.9	54.6	53.2	147	146.0
TRIA (50 ml ha <sup>-1</sup> )	3.6	4.1	5.7	6.4	75.2	88.7	185.3	190.6	50.5	49.3	146	144.0
NAA (10 ppm)	4.4	5.0	6.5	7.6	83.7	100.2	190.4	200.4	52.7	51.8	147	145.0
Control	3.8	4.0	5.5	6.1	60.2	74.0	170.3	183.5	49.3	49.0	144	141.0
SEM ( $\pm$ )	<b>0.08</b>	<b>0.06</b>	<b>0.33</b>	<b>0.34</b>	<b>5.73</b>	<b>6.5</b>	<b>5.21</b>	<b>5.14</b>	<b>0.02</b>	<b>0.06</b>	<b>0.03</b>	<b>0.04</b>
LSD (0.05)	NS	NS	1.0	1.0	17.2	19.5	15.6	15.4	NS	NS	NS	NS

NAA: Alpha naphthal acetic acid, Na-p-NP: Sodium para- nitrophenolate, NS: Not significant, TRIA: Triacontanol

**Table 2: Effect of treatments on yield attributes and fruit yield of rabi and kharif chilli (pooled of two-year data)**

Treatment	Fruits plant <sup>-1</sup>		Fruit length (cm)		Fruit width (cm)		Fruit weight (g)		Fruit yield (t ha <sup>-1</sup> )	
	Rabi	Kharif	Rabi	Kharif	Rabi	Kharif	Rabi	Kharif	Rabi	Kharif
Na-p-NP (4 ml L <sup>-1</sup> )	70.5	50.7	4.2	4.4	1.30	1.32	3.58	3.65	9.30	6.91
Na-p-NP(8 ml L <sup>-1</sup> )	82.0	68.5	4.5	4.7	1.51	1.52	3.92	4.00	11.82	10.84
Na-p-NP(16 ml L <sup>-1</sup> )	108.2	92.0	4.9	5.2	1.61	1.65	4.10	4.20	15.60	14.10
TRIA (50 ml ha <sup>-1</sup> )	60.8	42.7	4.0	4.3	1.36	1.40	3.43	3.50	7.56	5.80
NAA (10 ppm)	65.2	45.8	4.1	4.5	1.42	1.45	3.54	3.60	8.49	6.35
Control	54.6	37.5	3.4	3.7	1.20	1.25	3.05	3.10	6.10	4.50
SEM ( $\pm$ )	<b>2.87</b>	<b>2.70</b>	<b>0.21</b>	<b>0.24</b>	<b>0.04</b>	<b>0.03</b>	<b>0.07</b>	<b>0.07</b>	<b>0.48</b>	<b>0.43</b>
LSD (0.05)	<b>8.60</b>	<b>8.10</b>	<b>0.60</b>	<b>0.70</b>	<b>0.11</b>	<b>0.10</b>	<b>0.20</b>	<b>0.21</b>	<b>1.41</b>	<b>1.27</b>

NAA: Alpha naphthal acetic acid, Na-p-NP: Sodium para- nitrophenolate, TRIA: Triacontanol

of phenolic chemicals with gibberellins, which stimulate cell elongation (Taiz and Zeiger, 2002).

#### Number of flowers and fruits plant<sup>-1</sup>

As the number of flowers are directly related to the number of fruits of the plant, it affects the the chilli crop yield. When all PGRs were applied, the numbers of flowers and fruits produced by plant<sup>-1</sup> rose dramatically when compared to the untreated control (Table 1 and 2). The highest number of flowers plant<sup>-1</sup> was observed with the application of Na-p-NP at 16 ml L<sup>-1</sup> (238.2 and 276.9), whereas it was the lowest in control treatment (170.3 and 183.5) during *rabi* and *kharif* seasons, respectively. Many studies reveal that application of PGRs decreases flower and fruit drops, thus increasing the chilli yield (Mahindre *et al.*, 2018; Anolisa *et al.*, 2020). Similarly, the highest number of fruits plant<sup>-1</sup> was observed in Na-p-NP at 16 ml L<sup>-1</sup> with about 98.17 and 145.33% increase over control treatment in *rabi* and *kharif* seasons, respectively. These findings were in agreement with Chaudhary *et al.* (2006), Sarker *et al.* (2009), Das *et al.* (2015), Chouhan *et al.* (2017) and Kesumawati *et al.* (2019).

#### Days to 50% flowering and maturity

There was no significant effect of PGRs on the days to 50% flowering and maturity in both the seasons (Table 1). Maximum number of days (54.6 DAT) was required for 50% flowering in Na-p-NP at 16 ml L<sup>-1</sup> in *rabi* chilli whereas, it was attained at 53.2 DAT in *kharif*. In contrast, the untreated control treatment took minimum time (49.3 DAT for *rabi* and 49.0 DAT for *kharif*) to attain 50% flowering. Similarly, the plots treated with Na-p-NP at 16 ml L<sup>-1</sup> took maximum number of days (146 and 147 days in *rabi* and *kharif*, respectively) to mature, and the crop plants in untreated control attained maturity faster. This might be due to the enhanced vegetative growth caused by application of PGRs in early vegetative stage, and also PGRs could induce greenness for longer time due to higher photosynthetic activity. However, many contrasting results were reported on early flowering with the application of various PGRs (Kannan *et al.*, 2009; Tapdiya *et al.*, 2018).

#### Fruit length, width and weight

Data on fruit length of chilli indicated significant variation among different treatments (Table 2).

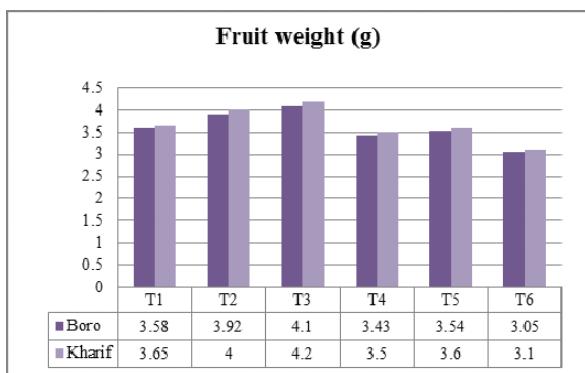


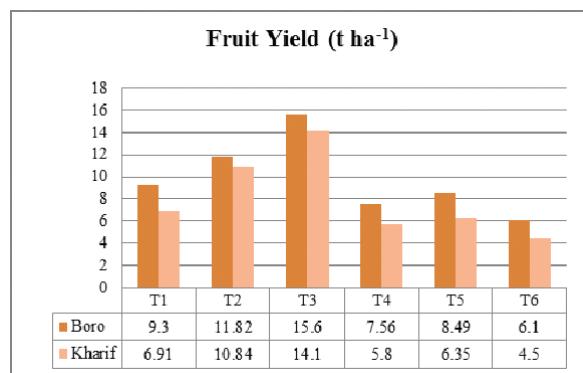
Fig. 1 : Effects of PGRs on fruit weight (g) in both seasons

Maximum fruit length was observed with the application of Na-p-NP at 16 ml L<sup>-1</sup> in both *rabi* (4.9 cm) and *kharif* (5.2 cm) seasons, whereas it was the lowest in untreated control. This might be affected by the availability of growth regulators within plants, causing cell expansion and elongation (Singh *et al.*, 1994). The fruit width also varied significantly with the application of PGRs (Table 2). Use of Na-p-NP at 16 ml L<sup>-1</sup> recorded the highest fruit width (1.61 cm in *rabi* and 1.65 cm in *kharif*). The lowest fruit width was recorded in untreated control (1.20 cm in *rabi* and 1.25 cm in *kharif*). This might be due to the fact that the plants' increased physiological activity and vegetative growth caused a buildup of assimilates (Joshi and Singh, 2010). These results were in consonance with the findings of Sharma *et al.* (1999), Tapdiya *et al.* (2018) and Pundir *et al.* (2020).

Application of Na-p-NP at 16 ml L<sup>-1</sup> recorded the highest fruit weight (4.10 g in *rabi* and 4.20 g in *kharif*) (Fig. 1), being 34.43 and 35.48% higher than the untreated control, respectively. Fruit width, length and number of seeds fruit<sup>-1</sup> altogether had a positive impact on increased fruit weight with the use of Na-p-NP at 16 ml L<sup>-1</sup>. These results were in conformity with the findings of Tapdiya *et al.* (2018).

#### Fruit yield

All the PGRs exhibited a significant influence on fruit yield. Use of Na-p-NP at 16 ml L<sup>-1</sup> recorded the highest fruit yield (15.60 and 14.10 t ha<sup>-1</sup>), whereas the untreated control recorded the lowest fruit yield (6.10 and 4.50 t ha<sup>-1</sup>) in *rabi* and *kharif* chillies (Table 2). Compared with the untreated control, there was a significant increase in fruit yield (155.74 and 213.33%) with the application of Na-p-NP at 16 ml L<sup>-1</sup>, whereas it was the lowest with the use of TRIA 250 ml ha<sup>-1</sup> (23.93 and 28.89%) in *rabi* and *kharif* seasons, respectively (Fig. 2). The plants treated with PGRs produced the highest fruit yield (Balraj *et al.*, 2002; Vandana and

Fig. 2 : Effects of PGRs on fruit yield (t ha<sup>-1</sup>) in both seasons

Verma, 2014). Photosynthetic activity of plants might be increased with the use of PGRs, resulting in the increased synthesis of carbohydrates and associated products, which led to the increased fruit size and weight, and ultimately higher productivity. These results were in consonance with the earlier findings of Chaudhary *et al.* (2006), Patel *et al.* (2016), Tapdiya *et al.* (2018), Mahindre *et al.* (2018).

As revealed from the present findings, three rounds of foliar spray of Na-p-NP at 16 ml L<sup>-1</sup> (25, 45-50 and 65-70 DAT) might be recommended for improving the growth, yield attributes and fruit yield of *rabi* and *kharif* chillies.

#### REFERENCES

- Ahmed, I.H., Ali, E.F., Gad, A.A., Bardisi, A., El-Tahan, A.M., Abd Esadek, O.A., El-Saadony, M.T. and Gendy, A.S. 2021. Impact of plant growth regulators spray on fruit quantity and quality of pepper (*Capsicum annuum* L.) cultivars grown under plastic tunnels. *Saudi J. Bio. Sci.*, **29**: 2291-2298.
- Akhter, S., Mostarin, T., Khatun, K., Akhter, F. and Parvin, A. 2018. Effects of plant growth regulator on yield and economic benefit of sweet pepper (*Capsicum annuum* L.). *Agric.*, **16**(2): 58-64.
- Anolisa, M., Al-Imran, R.H., ATM, R.I. and Subroto, K.D. 2020. Effect of plant growth regulators on growth and yield of chili (*Capsicum annuum* L.). *J. Phyto.*, **12**: 117-120.
- Anonymous. 2021a. CHILLI OUTLOOK REPORT – January to May 2021. Agricultural Market Intelligence Centre, ANGRAU, Lam. Pp.1. <https://angrau.ac.in/downloads/AMIC/CHILLI%20OUTLOOK%20REPORT%20-%20January%20to%20May%202021.pdf>
- Anonymous. 2021b. Chilli Outlook - August 2021. Agricultural Market Intelligence Centre, PJTSAU. Pp.1.<https://pjtsau.edu.in/files/AgriMkt/2021/August/chilli-August-2021.pdf>

## *Effect of Sodium Para Nitrophenolate 0.3% SL on Growth and Yield of Chilli*

- Balraj, R., Kudikeri, M. B. and Revanappa. 2002. Effect of growth regulators on growth and yield of chilli (*Capsicum annuum L.*) at different pickings. *Indian J. Hort.*, **59**: 84-88.
- Chaudhary, B.R., Sharma, M.D., Shakya, S.M. and Gautam, D.M. 2006. Effect of plant growth regulators on growth, yield and quality of chilli (*Capsicum annuum L.*). *J. of Agril. Res.*, **27**: 65-68.
- Chouhan, K.S., Baghel, S.S., Mishra, K., Singh, A.K. and Singh, V. 2017. Effect of varieties and integrated nutrient management on growth and yield of chilli (*Capsicum annuum L.*). *Int. J. Pure App. Biosci.* **5**(4): 2114-2120.
- Das, S.K., Sarkar, M.D., Alam, M.J., Robbani, M.G. and Kabir, M.H. 2015. Influence of plant growth regulators on yield contributing characters and yield of bell pepper (*Capsicum annum*) varieties. *J. Plant Sci.* **10**(2): 63-69.
- Deshmukh, D.A., Telang, S.M. and Patil, S.S. 2010. Influence of foliar application of growth hormones and fertilizers on the field and ascorbic content in chilli cv Parbhani Tajas. *Asian J. Soil Sci.* **5**(1): 114-115.
- Djanaguiraman, M., Sheeba, J.A., Devi, D.D. and Bangarusamy, U. 2005. Effect of Atonik seed treatment on seedling physiology of cotton and tomato. *J. Biol. Sci.*, **5**: 163-169.
- dos Anjos, G.L., Moreira, G.C., Carneiro, C.N. and Dias, F.D.S. 2022. Effect of phytochemicals on the composition of phenolic compounds in chili peppers (*Capsicum frutescens*) and exploratory analysis. *Sci. Hortic.*, **292**: 110660.
- Emongor, V. 2007. Gibberellic Acid (GA<sub>3</sub>) Influence on vegetative growth, nodulation and yield of cowpea (*Vigna unguiculata L.*) Walp. *J. Agron.*, **6**: 509-517.
- Gare, B.N., Raundal, P.U. and Burli, A.V. 2017. Effect of plant growth regulators on growth, yield and yield attributing characters of rainfed chilli (*Capsicum annuum L.*). *Adv. Agril. Res. Tech.*, **1**(2): 195-197.
- Gianfagna, T. 1995. Natural and synthetic growth regulators and their use in horticultural and agronomic crops. In Plant hormones. Springer, Dordrecht. Pp. 751-773.
- Joshi, C. and Singh, D.K. 2010. Effect of plant growth regulators on chilli. *J. Vegetables Sci.*, **28**(1): 23-72.
- Joshi, N. C., Singh, D. K. and Jain, S. K. 1999. Response of plant bioregulators on growth and yield of chilli during summer season. *Adv. Hort. and For.*, **7**: 95-99.
- Kalshyam, M. K., Kumar, J., Mohan, B., Singh, J.P., Ram, N. and Rajbeer. 2011. Effect of plant growth hormone and fertilizer on growth and yield parameters in chilli (*Capsicum annum L.*) cv. Pusa Jwala. *Asian J. Hort.*, **6**(2): 316-318.
- Kannan, K., Jawaharlal, M. and Prabhu, M. 2009. Effect of plant growth regulators on growth and yield parameters of Paprika cv. ktpl- 19. *Agric. Sci. Digest*, **29**(3): 157-162.
- Kar, B., Patra, C., Padhiary, A.K. and Mohanty, S. E. 2016. Effect of different levels of potassium and plant growth regulators on different yield attributes of chilli (*Capsicum annuum L.*). *Int. J. Sci. Env. Tech.*, **5**(5): 3147-3159.
- Kariali, E. and Mohapatra, P. K. 2007. Hormonal regulation of tiller dynamics in differentially-tillering rice cultivars. *Pl. Growth Regulation*, **53**(3): 215-223.
- Kesumawati, E., Apriyatna, D. and Rahmawati, M. 2019. The effect of shading levels and varieties on the growth and yield of chili plants (*Capsicum annum L.*). *IOP Conf. Ser. Earth Environ. Sci.* **425**(1): 012080.
- Kozak, M., Malarz, W., Serafin-Andrzejewska, M. and Kotecki, A. 2008. The effect of different sowing rate and Asahi SL treatment on soybean sowing value. In: *Monographs Series: Biostimulators in Modern Agriculture: Field Crops*, ed: Z.T.D<sup>1</sup> browski, Warsaw, Editorial House Wieso. Pp. 85-91.
- Mahindre, P.B., Jawarkar, A.K., Ghawade, S.M. and Tayade, V.D. 2018. Effect of different concentration of plant growth regulators on growth and quality of green chilli, *JPP*, SP **1**: 3040-3042.
- Patel, V.P., Plal, E. and John, S. 2016. Comparative study of the effect of plant growth regulators on growth, yield and physiological attributes of chilli (*Capsicum annum L.*) cv. Kashi Anmol. *Int. J. Farm Sci.* **6**(1): 199-204.
- Pundir, D., Singh, S. and Saxena, A.K. 2020. Response of plant growth regulators (NAA and GA3) on growth and yield attributes of chilli (*Capsicum annuum L.*) at Dehradun valley region. *Int. J. Chem. Stud.*, **8**(5): 556-559. DOI: 10.22271/chemi.2020.v8.i5h.10354
- Sarker, P., Hossain, T., Mia, M.A., Islam, R. and Miah, M.N.A. 2009. Effect of NAA on growth, yield and quality of chilli (*Capsicum frutescens*). *Bangladesh Res. Pub. J.* **2**(3): 612-617.
- Shah, S.H. and Ahmad, I. 2006. Effect of gibberellic acid spray on growth, nutrient uptake and yield attributes during various growth stages of black cumin (*Nigella sativa L.*). *Asian J. Pl. Sci.*, **5**: 881-884.

- Sharma, A.K., Ratian, R.S. and Pathania, N.K. 1999. Effect of plant growth regulators on yield and morphological traits in brinjal (*Solanum melongena* L.). *Agric. Sci. Dig.*, **12**: 219-222.
- Singh, D.K., Lal, G. and Singh, R.P. 1994. Effect of plant growth regulators on the fruit set, yield and quality of chilli (*Capsicum annuum* L.) cultivars. *Adv. Hortic. Forest.*, **4**:133-141.
- Sreenivas, M., Sharangi, A.B. and Raj, A.C., 2017. Evaluation of bio-efficacy and phytotoxicity of gibberellic acid on chilli. *J. Crop and Weed.*, **13**(3): 174-177.
- Srivastava, M. 2007. Effect of plant bioregulators on growth and yield attributing characters in chilli (*Capsicum annuum* L.) cv. Jawahar Mirchi, M.Sc (Agri). Thesis, JNKVV Collage of Agriculture, Jabalpur, Madhya Pradesh.
- Surendar, P., Sekar, K., Sha, K. and Kannan, R. 2020. Effect of plant growth regulators on growth of chilli (*Capsicum annuum* L.). *Plant Arch.*, **20**(1): 1544-1546.
- Taiz, L. and Zeiger, E. 2002. *Plant Physiology*. Sunderland: Sinauer Associates, Inc., Publishers.
- Tamilselvi, C. and Vijayaraghavan, H. 2014. Impact of plant growth regulators and formulations on growth of chilli (*Capsicum annuum* L.). *J. Plant Gene and Trait*, **5**: 1-3.
- Tapdiya, G.H., Gawande, P.P., Ulemale, P.H., Patil, R.K. and Naware, M.S. 2018. Effect of growth regulators on quantitative characters of chilli (*Capsicum annuum* L.). *Int. J. Curr. Microbiol. App. Sci.*, **6**: 2151–2157.
- Tiwari, D.K., Pandey, P., Giri, S.P. and Dwivedi, J.L. 2011. Effect of GA3 and other plant growth regulators on hybrid rice seed production. *Asian J. Plant Sci.*, **10**(2): 133-139.
- Valero, D., Zapata, P.J., Martinez-Romero, D., Guillén, F., Castillo, S. and Serrano, M. 2014. Pre-harvest treatments of pepper plants with nitrophenolates increase crop yield and enhance nutritive and bioactive compounds in fruits at harvest and during storage. *Food Sci. Tech. Int.*, **20**(4): 265-274.
- Vandana, P. and Verma, L.R. 2014. Effect of spray treatment of pant growth substances at different stages on growth and yield of sweet pepper (*Capsicum annum* L.) cv. Indra. *Int. J. Agril. Res.*, **2**: 235-240.
- Vega-Alfaro, A., Ramírez-Vargas, C., Chávez, G., Lacayo, F., Bethke, P.C. and Nienhuis, J. 2021. Flowering time and productivity of interspecific grafts between pepper species in contrasting high tunnel-sheltered and open-field production environments in Costa Rica. *Hort. Tech.*, **1** (aop): 1–10.